

## 510(k) Summary

Submitter:

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Date Summary

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Device Name:

Mouse Anti-Human T-cell, CD4/FITC, MT310+ Mouse Anti-Human T-cell, CD8/RPE, DK25

Device

Classification:

Class II according to 21 CFR 864.5220, on the basis that monoclonal antibodies

are accessories for automated differential cell counters.

Panel:

This device classification is uncer the Hematology and Pathology devices panel,

Division of Clinical Laboratory Cevices.

Product Code:

GKZ

Predicate Device(s):

CytoStat/Coulter Clone T8-RD1 T4-FITC

Device Description:

Purified mouse anti-human CD4, CloneMT310, conjugated with fluorescein isothiocyanate, isomer 1 (FITC) + purified mouse anti-human CD8, Clone DK25, conjugated with R-phycoerythrin, present in 0.05M Tris-HCl buffer, pH 7.2, 15

mM NaN<sub>3</sub>, 0.1M NaCl, stabilized with 1% carrier protein

Subpopulations of lymphocytes may be stained with fluorochrome-conjugated antibody and evaluated in perioheral blood specimens when contaminating red blood cells (RBC's) are lysed prior to flow cytometric analysis. A subpopulation

of WBC's are selected for assessment based upon cell morphology.

Intended Use:

For In Vitro Diagnostic Use

Mouse Anti-Human T-cell, CD4/FITC, MT310 + Mouse Anti-Human T-cell, CD8/RPE, DK25 (DAKO CD4/CD8) has been developed for use in flow cytometry for the analysis of T-helper/inducer cells and T-cytotoxic/suppressor This reagent allows simultaneous detection and quantification of helper/inducer T-cells (CD4 + cells) and cytotoxic/supprossor T-cells (CD8 + cells) in peripheral blood of normal and pathological conditions such as immunodeficiency disorders. It is one component of the suggested monoclonal antibody (MAb) combinations for routine immunophenotyping of lymphocytes in peripheral blood using flow cytometry.

Comparison of Technological Characteristics

Performance characteristics have been established by clinical evaluation of compared to the individual single reagent predicate devices that quantitatively measure CD4<sup>+</sup> and CD8<sup>+</sup> T-cells that have been previously cleared by FDA (CytoStat/CoulterClone T8-RD-1/T4-FITC). When flow cytometric tests of peripheral blood samples obtained from apparently healthy adults were completed, correlation of CytoStat/CoulterClone T8-RD-1/T4-FITC with DAKO CD4/CD8 approached a direct 1 : 1 comparison for measurement of CD4+ cells. Correlation of CytoStat/CoulterClone T8-RD-1/T4-FITC with DAKO CD4/CD8 approached a direct 1: 1 comparison for measurement of CD8+cells. Data for the measurement of CD4+ T-cells by DAKO CD4/CD8 reagent compared to CytoStat/CoulterClone T8-RD-1/T4-FITC on peripheral blood samples obtained from apparently healthy adults as well as ill patients gave a correlation greater than 0.98 using the whole blood method for flow cytometry. Data for the measurement of CD8+ T-cells by DAKO CD4/CD8 reagent compared to CytoStat/CoulterClone T8-RD-1/T4-FITC gave a correlation greater than 0.98 using the whole blook method for flow cytometry.

The CD4 antibody clone, MT31(), was clustered at the Second Leukocyte Typing Workshop, Boston, 1984. The CD8 antibody clone, DK25, was clustered at the Third Leukocyte Typing Workshop, Oxford, England, 1986 under another clone designation.

Linearity testing of DAKO CD4/FITC using CEM cells gave the following linear equation:

$$y = 4.26\% + 0.93x$$
;  $r = 0.999$ 

Linearity testing of DAKO CD8/FPE using JM cells gave the following linear equation:

$$y = 0.06\% + 1.01x$$
;  $r = 0.999$ 

In addition, reproducibility of DAKO reagents using replicates (from peripheral blood) run on two different flow cytometers was measured at three concentrations of each antigen. Cross-reactivity of CD4/CD8 with peripheral blood cells (red blood cells, monocytes, granulocytes, lymphocytes, and platelets) was measured.

Conclusions:

Results of the above testing as well as the information provided by the Second and Third Leukocyte Typing Workshops indicate that the CD4/CD8 reagent performs as well as CytoStat/CoulterClone T8-RD-1/T4-FITC in the detection and enumeration of CD4<sup>+</sup> and CD8<sup>+</sup> lymphocytes using flow cytometry. Safety of the DAKO CD4/CD8 reagent and its predicate device is high as are all reagents used for in vitro testing.

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